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Tetrahedron

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Synthesis of 2-amino-1,3-diols incorporating the cyclobutane ring

Marta Pérez-Fernández, Alberto Avenoza, Jesús H. Busto*, Jesús M. Peregrina*, Fernando Rodríguez

Departamento de Química, Universidad de La Rioja, Grupo de Síntesis Química de La Rioja, U.A.-C.S.I.C., E-26006 Logroño, Spain

ARTICLE INFO

Article history: Received 14 April 2008 Received in revised form 1 July 2008 Accepted 3 July 2008 Available online 8 July 2008

ABSTRACT

The synthesis of free and protected 2-amino-1,3-diols with threoninol substructure that incorporate a conformational restriction defined by the cyclobutane ring is reported. The key step in the synthesis of these target compounds, namely *cis*- and *trans*-c₄-threoninols, is the addition of methylmagnesium bromide to a cyclobutanone derivative. The selectivity of the reaction is modulated by the solvent. © 2008 Elsevier Ltd. All rights reserved.

1. Introduction

In the last few years, the synthesis of cyclobutane compounds has received great attention, due to their use as building blocks for the preparation of natural products. Moreover, the development of new synthetic strategies has allowed an easy access to this kind of rings.¹ In this context, a number of cyclobutane-containing derivatives such as cyclobutane-constrained α -amino acids,^{2a,b} cyclobutane-constrained β -amino acids,^{2c} cyclobutane peptide-turned mimics^{2d} or cyclobutane nucleoside analogues^{2e} have been reported recently in the literature.

Threoninol, a 2-amino-1,3-diol derivative, is an important compound obtained by reduction of the natural amino acid threonine (Thr) (Fig. 1). It has been proved to have some interesting applications. Thus, some conveniently protected derivatives could be used as analogues of ribonucleotides and modification of their structure would provide a library of compounds.³ In nanomaterial chemistry, both the enantiomers of threoninol were used as linkers between DNA chains. The use of each enantiomer afforded the corresponding handed helix of DNA clusters.⁴ On the other hand, somatostatin is a tetradecapeptide involved in very important biological processes. The receptors of this peptide are found to be expressed in 90% of carcinoid tumours. In terms of activity, octreotide is a somatostatin analogue more effective than somatostatin itself, due to a longer half life in vivo. Remarkably, threoninol is present in octreotide along with other p-amino acids.⁵

We intend to focus the attention on the 2-amino-1,3-diols and the development of new related structures. As part of our research program,^{6,7} we have synthesized different cyclobutane derivatives,^{6a,b} and wish to extend this work to the preparation of restricted 2-amino-1,3-diols containing the cyclobutane substructure. Particularly, it is our purpose to develop a new route for



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Figure 1. Threoninols and their cyclobutane derivatives.

the synthesis of cis- and trans-stereoisomers of 2-amino-2-(hydroxymethyl)-1-methylcyclobutanol, abbreviated as c₄-threoninol and c₄-allo-threoninol, trans-1 and cis-1, respectively (Fig. 1).

The synthetic methodology that allows an easy formation of the cyclobutane ring involves the reaction of methyl 2-acetamidoacrylate **2** with ketene diethyl acetal **3** in a formal [2+2] cycloaddition. In this way, a tandem Michael–Dieckmann-type process gives compound **4**, which is then transformed into the versatile intermediate **5** (Scheme 1).⁷ We have planned to obtain c_4 -threoninols from **5**, by using methylmagnesium bromide to incorporate the methyl group into the position 2 of the cyclobutane ring (Scheme 2).

2. Results and discussion

As planned, the cyclobutane derivatives with threoninol substructure were easily achieved from **5**, by reaction with methylmagnesium bromide in different solvents and at different temperatures. This kind of reaction has been extensively studied.⁸



^{*} Corresponding authors. Fax: +34 941299621.

E-mail addresses: hector.busto@unirioja.es (J.H. Busto), jesusmanuel.peregrina@ unirioja.es (J.M. Peregrina).

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Scheme 1. Starting material for the preparation of c₄-threoninol derivatives (TBDPS=tertbutyldiphenylsilyl).



Scheme 2. Addition of methylmagnesium bromide to compound 5.

More particularly, Pérez-Ossorio et al. described the addition of phenylmagnesium bromide to 3-phenylbutanone in different solvents at 30 °C.^{8d} A linear correlation between stereoselectivity and the $E_{\rm T}(30)$ parameter of the solvent was found. Similarly, the same trend could be identified in the experiments carried out under our conditions (Scheme 2 and Table 1). Thus, when the addition of methylmagnesium bromide to **5** takes place in THF, dimethoxy-ethane (DME) or dioxane, the stereoisomer *trans*-**6** is obtained as the major compound, whereas in diethyl ether (Et₂O) or hexane, solvents with lower $E_{\rm T}(30)$ values, the stereoisomer *cis*-**6** was the main product (Table 1). It is worth to mention that, to the best of our knowledge, studies correlating solvent properties and stereo-selectivity of this kind of reaction are not very common in the literature.

On the basis of the data gathered in Table 1, it can be deduced that the selectivity in this reaction seems to be controlled mainly by the solvent, since the change of temperature is not very significant in general. In fact, the most important effect was observed when the reaction was refluxed in dioxane, the solvent with the highest boiling point (Table 1, entries 7 and 8). The best selectivity in favour of *cis*-**6** was achieved with Et₂O as solvent, at 30 and $-78 \degree C$, ⁹ however, the conversion at the latter temperature is slightly lower (Table 1, entries 9 and 11). Regarding *trans*-**6**, the best results concerning selectivity were observed in DME at 30 °C (Table 1, entry 5). Nonetheless, there was just a 50% conversion. When the reaction was conducted in the same solvent at 85 °C, the conversion

Table 1

Conditions and results for the addition of MeMgBr to cyclobutanone **5** as depicted in Scheme 2

Entry	Solvent	T (°C)	Conversion ^a	Yield % ^b	cis- 6 /trans- 6 ratio ^c
1	THF	-78	25		24/76
2	THF	0	100	77	19/81
3	THF	30	100	80	22/78
4	THF	66	100		26/74
5	DME	30	50		10/90
6	DME	85	100	52 ^d	18/82
7	Dioxane	30	60		31/69
8	Dioxane	101	100	41 ^d	50/50
9	Et ₂ O	-78	90		70/30
10	Et ₂ O	0	100	86	62/38
11	Et ₂ O	30	100	90	70/30
12	Hexane	30	75		61/39
13	Hexane	69	75		57/43

^a Determined by ¹H NMR and/or HPLC.

^b Yield after column chromatography.

^c See Ref. 9.

^d At these high temperatures, the yield decreased due to the formation of several unidentified sideproducts.

reached 100% (Table 1, entry 6). Unfortunately, the selectivity decreased slightly and a more complex mixture of products was observed. The best option for the synthesis of *trans*-**6** considering both, conversion and selectivity, is to carry out the reaction in THF at 0 °C (Table 1, entry 2). In these conditions, this isomer was obtained in a good overall yield after column chromatography.

As previously mentioned, similar results to those obtained in THF, DME or dioxane have been reported in the literature for other ketones.^{8d-f} As expected, when these solvents were used, *trans*-**6** was the major product⁹ (Table 1). This result can also be explained if the organomagnesium compound attacks following the mechanism of the Felkin–Ahn model,¹⁰ in which, the bulky substituent is located almost orthogonal to the carbonyl group, allowing the nucleophile to attack *anti* to this substituent and avoid steric repulsions. A graphic representation is shown in Figure 2.

In contrast, the use of Et_2O or hexane led to the other stereoisomer, *cis*-**6**, as the major product.⁹ This change of selectivity can also be understood taking into account the different behaviour of the organomagnesium compound in solvents like Et_2O or hexane that might favour formation of the chelating intermediate represented in Figure 2. In this case, the addition probably follows the chelation model,⁸ where the nucleophilic attack takes place from the opposite face, and therefore the other isomer is obtained.

Compounds *cis*-**6** and *trans*-**6** were isolated after column chromatography and their stereochemistry was assigned by 2D NOESY experiments. These experiments were performed using phasesensitive ge-2D NOESY¹¹ in CDCl₃ and showed, as main facts, positive cross-peaks between the NHAc and CH₃ signals for compound *trans*-**6**, and between CH₂OTBDPS and CH₃ for compound *cis*-**6**. The absence of cross-peaks between the NHAc and CH₃ signals confirms the relative configuration of the latter one (Fig. 3).

After isolation of *cis*-**6** and *trans*-**6**, simple transformations of the functional groups are required to achieve the c_4 -threoninols. All the protected groups in both the compounds were removed by hydrolysis in a 3 N HCl aqueous solution at 70 °C, to give the hydrochloride salts of *cis*-**1** and *trans*-**1**, respectively, in good yields (Scheme 3).

c₄-Threoninols with different protection patterns are necessary for synthetic purposes. In this context, the orthogonal protection of derivatives **6** was achieved by introduction of the methoxymethyl (MOM) group in the secondary alcohol. Thus, treatment of *cis*-**6** and *trans*-**6** with methoxymethyl chloride (MOMCl) and diisopropylethylamine (DIEA) led to *cis*-**7** and *trans*-**7**, respectively, in good overall yields (Scheme 4).

To obtain derivatives with the two alcohol groups free, the primary alcohol functions of *cis*-**6** and *trans*-**6** (protected with TBDPS) were reacted with tetrabutyl ammonium fluoride (TBAF) in THF at room temperature. In this way *cis*-**8** and *trans*-**8** were cleanly obtained (Scheme 4).





Figure 3. 2D-NOESY experiments of *trans*-6 (up) and *cis*-6 (bottom).



Scheme 3. Synthesis of c4-threoninols cis-1 and trans-1.



Scheme 4. Synthesis of differently protected c₄-threoninols.

Finally, to synthesize the primary alcohol protected c₄-threoninols *cis*-**9** and *trans*-**9**, their corresponding *cis*-**7** and *trans*-**7** precursors were subjected to the silyl group cleavage using again TBAF in THF (Scheme 4).

3. Conclusions

In summary, we have developed a synthesis of new 2-amino-1,3-diols with cyclobutane structure in different protected and unprotected forms, to be used in the synthesis of other interesting compounds. The key step, which can be modulated by the solvent, was the addition of methylmagnesium bromide to the corresponding cyclobutanone derivative, allowing to obtain the required products in good overall yields and selectivities.

4. Experimental

4.1. General procedures

Solvents were purified according to standard procedures. Analytical TLC was performed using Polychrom SI F₂₅₄ plates. Column chromatography was performed using silica gel 60 (230-400 mesh). ¹H and ¹³C NMR spectra were recorded on a Bruker ARX-300 spectrometer using CDCl₃ with TMS as the internal standard or using CD₃OD or D₂O with TMS as the external standard with a coaxial microtube (chemical shifts are reported in parts per million on the δ scale, coupling constants in Hz). The assignment of all separate signals in the ¹H NMR spectra was made on the basis of coupling constants, ge-COSY experiments and ge-HSQC experiments on a Bruker AVANCE 400 spectrometer. This spectrometer was also used for 2D NOESY experiments described in the text. These experiments were processed with Mestre Nova software (Mestrelab Research, Spain). Melting points were determined on a Büchi SMP-20 melting point apparatus and are uncorrected. Microanalyses were carried out on a CE Instruments EA-1110 analyser and are in good agreement with the calculated values.

4.1.1. trans-N-(2-Hydroxy-1-(tert-butyldiphenylsilyloxymethyl)-2-methylcyclobutyl)acetamide (trans-**6**)

Compound **5** (1 g, 2.5 mmol) was solved in dry THF (50 mL) and was cooled to 0 °C. Then, 2.5 mL of MeMgBr (3 M in THF) was slowly added. The resulting solution was stirred for 14 h at this

temperature. A saturated NH₄Cl solution was added (20 mL) and the aqueous phase was washed with ethyl acetate $(3 \times 30 \text{ mL})$. The organic phase was dried, filtered, and concentrated in vacuo to give a mixture of compounds cis-6 and trans-6 in an 81:19 ratio in favour of compound trans-6. The mixture was separated by column chromatography (hexane/ethyl acetate, 1:1) to give 625 mg of compound *trans*-**6** and 175 mg of compound *cis*-**6**, both as white solids (800 mg, 77%). Compound *trans*-**6**. Mp: 138–140 °C. ¹H NMR (CD₃OD) δ 1.07 (s, 9H, C(CH₃)₃), 1.32 (s, 3H, CH₃), 1.43–1.51 (m, 1H, H₃), 1.78-1.83 (m, 1H, H₄), 1.87 (s, 3H, COCH₃), 1.94-2.03 (m, 1H, H₄), 2.10-2.16 (m, 1H, H₃), 3.06 (s, 1H, OH), 3.98 (d, 1H, *I*=10.8, CH₂O), 4.25 (dd, 1H, J=2.0, J=10.8, CH₂O), 5.73 (br s, 1H, NH), 7.36-7.43 (m, 6H, Ph), 7.60–7.65 (m, 4H, Ph). ¹³C NMR (CD₃OD) δ 19.4 (C(CH₃)₃), 22.1 (C₄), 23.0 (CH₃CO), 24.2 (CH₃), 27.0 (C(CH₃)₃), 30.7 (C₃), 63.4 (CH₂O), 64.0 (C₁), 74.7 (C₂), 127.7, 129.7, 133.5, 135.5 (Ph), 170.3 (CO). ESI⁺ (m/z)=412.7. Anal. Calcd for C₂₄H₃₃NO₃Si: C, 70.03; H, 8.08; N, 3.40. Found: C, 70.15; H, 8.10; N, 3.45.

4.1.2. cis-N-(2-Hydroxy-1-(tert-butyldiphenylsilyloxymethyl)-2methylcyclobutyl)acetamide (cis-**6**)

Compound 5 (1 g, 2.5 mmol) was solved in dry Et₂O (50 mL) at 30 °C. Then, 2.5 mL of MeMgBr (3 M in Et₂O) was slowly added. The resulting solution was stirred for 14 h at this temperature. A saturated NH₄Cl solution was added (20 mL) and the aqueous phase was washed with ethyl acetate $(3 \times 30 \text{ mL})$. The organic phase was dried, filtered and concentrated in vacuo to give a mixture of compounds cis-6 and trans-6 in a 70:30 ratio in favour of compound cis-6. The mixture was subjected to column chromatography (hexane/ethyl acetate, 1:1) to give 656 mg of compound cis-6 and 282 mg of compound trans-6, both as white solids (938 mg, 90%). Compound *cis*-**6**. Mp: 144–146 °C. ¹H NMR (CDCl₃) δ 1.07 (s, 9H, C(CH₃)₃), 1.40 (s, 3H, CH₃), 1.67-1.75 (m, 1H, H₄), 1.79-1.87 (m, 1H, H₃), 1.94 (s, 3H, COCH₃), 2.05–2.12 (m, 1H, H₃), 2.25–2.32 (m, 1H, H₄), 2.98 (br s, 1H, OH), 3.81 (d, 1H, J=10.4, CH₂O), 3.94 (d, 1H, J=10.4, CH₂O), 6.22 (br s, 1H, NH), 7.37-7.43 (m, 6H, Ph), 7.61-7.64 (m, 4H, Ph). ¹³C NMR (CDCl₃) δ 19.3 (C(CH₃)₃), 23.1 (C₄), 23.7 (CH₃CO), 23.9 (CH₃), 26.9 (C(CH₃)₃), 32.1 (C₃), 64.1 (C₁), 64.8 (CH₂O), 76.1 (C₂), 127.8, 129.9, 133.2, 135.5 (Ph), 171.1 (CO). ESI⁺ (*m*/*z*)=412.7. Anal. Calcd for C₂₄H₃₃NO₃Si: C, 70.03; H, 8.08; N, 3.40. Found: C, 70.10; H, 8.12; N, 3.38.

4.1.3. trans-2-Amino-2-(hydroxymethyl)-1-methylcyclobutanol hydrochloride salt (trans-1·HCl)

The white solid *trans*-**6** (50 mg, 0.12 mmol) was suspended in an aqueous 3 N HCl solution (10 mL) and the mixture was heated at 70 °C. After stirring for 4 h, the solvent was evaporated in vacuo and the resulting residue was partitioned between H₂O and ethyl acetate. The aqueous phase was concentrated in vacuo to give the product *trans*-**1**·HCl (c₄-threoninol) as a pale oil (18 mg, 90%). ESI⁻ (*m*/*z*)=132.3. ¹H NMR (D₂O) δ 1.44 (s, 3H, CH₃), 2.07–2.13 (m, 4H, CH₂), 3.80–3.92 (m, 2H, CH₂OH). ¹³C NMR (D₂O) δ 24.6, 24.8, 33.4, 64.2, 66.1, 76.4. Anal. Calcd for C₆H₁₄ClNO₂: C, 42.99; H, 8.42; N, 8.36. Found: C, 43.01; H, 8.40; N, 8.29.

4.1.4. cis-2-Amino-2-(hydroxymethyl)-1-methylcyclobutanol hydrochloride salt (cis-**1**·HCl)

The white solid *cis*-**6** (50 mg, 0.12 mmol) was suspended in an aqueous 3 N HCl solution (10 mL) and the mixture was heated at 70 °C. After stirring for 3 h, the solvent was evaporated in vacuo and the resulting residue was partitioned between H₂O and ethyl acetate. The aqueous phase was concentrated in vacuo to give the product *cis*-**1**·HCl (c₄-*allo*-threoninol) as a pale oil (17 mg, 85%). ESI⁻ (*m*/*z*)=132.3. ¹H NMR (D₂O) δ 1.48 (s, 3H, CH₃), 1.73–1.89 (m, 1H, CH₂), 1.95–2.07 (m, 3H, CH₂), 3.92 (q, *J*=6.9, 2H, CH₂OH). ¹³C NMR (D₂O) δ 21.5, 22.8, 31.6, 62.2, 64.1, 74.5. Anal. Calcd for

C₆H₁₄ClNO₂: C, 42.99; H, 8.42; N, 8.36. Found: C, 42.98; H, 8.38; N, 8.31.

4.1.5. trans-N-(2-Methoxymethyloxy-1-(tert-butyldiphenylsilyloxymethyl)-2-methylcyclobutyl)acetamide (trans-7)

Compound trans-6 (200 mg, 0.49 mmol) was solved in DIEA (1.0 mL, 5.8 mmol), under argon atmosphere, at 0 °C. After that, MOMCl was added (0.37 mL, 4.86 mmol) and the mixture was maintained at this temperature for 1 h. The temperature was then increased to room temperature and the reaction was stirred for 14 h. The reaction mixture was diluted with ethyl acetate (15 mL) and washed with H₂O (2×20 mL) and brine (2×10 mL). The organic layer was dried, concentrated in vacuo and purified by silica gel column chromatography (hexane/ethyl acetate, 1:1) to give the product trans-7 (180 mg, 81%) as a colourless oil. ESI+ (m/z)=456.7. ¹H NMR (CDCl₃) δ 1.06 (s, 9H, C(CH₃)₃), 1.40 (s, 3H, CH₃), 1.80–1.85 (m, 2H, H₃, H₄), 1.92 (s, 3H, COCH₃), 1.96–2.12 (m, 2H, H₃, H₄), 3.25 (s, 3H, OCH₃), 3.88 (d, 1H, *I*=10.6, CH₂OSi), 4.33 (d, 1H, *J*=10.6, CH₂OSi), 4.54 (d, 1H, *J*=7.1, CH₂OCH₃), 4.78 (d, 1H, *I*=7.1, CH₂OCH₃), 5.55 (br s, 1H, NH), 7.36–7.44 (m, 6H, Ph), 7.65– 7.67 (m, 4H, Ph). ¹³C NMR (CDCl₃) δ 19.5 (C(CH₃)₃), 20.8 (CH₃), 22.6 (C₄), 23.9 (COCH₃), 27.0 (C(CH₃)₃), 30.0 (C₃), 55.3 (OCH₃), 63.2 (CH₂OSi), 64.6 (C₁), 80.4 (C₂), 92.2 (CH₂OCH₃), 127.7, 127.8, 129.7, 133.7, 133.8, 135.7 (Ph), 169.4 (CO). Anal. Calcd for C₂₆H₃₇NO₄Si: C, 68.53; H, 8.18; N, 3.07. Found: C, 68.56; H, 8.14; N, 3.03.

4.1.6. cis-N-(2-Methoxymethyloxy-1-(tert-butyldiphenylsilyloxymethyl)-2-methylcyclobutyl)acetamide (cis-**7**)

Compound cis-6 (260 mg, 0.63 mmol) was solved in DIEA (1.3 mL, 7.6 mmol), under argon atmosphere, at 0 °C. Then, MOMCl was added (0.48 mL, 6.3 mmol) and the mixture was maintained at this temperature for 1 h. The temperature was then increased to room temperature and the reaction was stirred for 14 h. The reaction mixture was diluted with ethyl acetate (15 mL) and washed with H_2O (2×20 mL) and brine (2×10 mL). The organic phase was dried and concentrated in vacuo and the residue was purified by silica gel column chromatography (hexane/ethyl acetate, 7:3) to give product cis-7 (220 mg, 77%) as a colourless oil. ESI⁺ (m/z)=456.7. ¹H NMR (CDCl₃) δ 1.07 (s, 9H, C(CH₃)₃), 1.50-1.57 (m, 4H, CH₃, H₄), 1.78 (t, 1H, J=8.7, H₃), 1.94 (s, 3H, COCH₃), 2.18-2.26 (m, 2H, H₃, H₄), 3.45 (s, 3H, OCH₃), 3.62 (d, 1H, *I*=10.8, CH₂OSi), 4.18 (d, 1H, *I*=10.8, CH₂OSi), 4.78 (d, 1H, *I*=6.7, CH₂OCH₃), 4.83 (d, 1H, *I*=6.7, CH₂OCH₃), 6.72 (br s, 1H, NH), 7.36-7.42 (m, 6H, Ph), 7.62–7.66 (m, 4H, Ph). ¹³C NMR (CDCl₃) δ 19.3 (C(CH₃)₃), 20.5 (CH₃), 20.8 (C₄), 24.1 (COCH₃), 26.9 (C(CH₃)₃), 31.0 (C₃), 55.6 (OCH₃), 63.1 (CH₂OSi), 64.0 (C₁), 78.7 (C₂), 92.0 (CH₂OCH₃), 127.5, 127.6, 129.6, 133.4, 133.6, 135.6 (Ph), 169.3 (CO). Anal. Calcd for C₂₆H₃₇NO₄Si: C, 68.53; H, 8.18; N, 3.07. Found: C, 68.52; H, 8.17; N, 3.02.

4.1.7. trans-N-(2-Hydroxy-1-(hydroxymethyl)-2-methyl-cyclobutyl)acetamide (trans-**8**)

To a solution of *trans*-**6** (390 mg, 0.95 mmol) in dry THF (20 mL), TBAF (0.95 mL, 1 M in THF) was added and the mixture was stirred for 1 h at room temperature. Then, the reaction was concentrated in vacuo and the residue was purified by silica gel column chromatography (MeOH/ethyl acetate, 5:95) to obtain the product *trans*-**8** as a white solid (120 mg, 75%). Mp: 130–132 °C. ¹H NMR (CD₃OD) δ 1.28 (s, 3H, CH₃), 1.54–1.57 (m, 1H, H₄), 1.77 (t, 1H, *J*=9.2, H₃), 1.91– 1.95 (m, 5H, COCH₃, H₃, H₄), 3.75 (d, 1H, *J*=11.6, CH₂O), 4.13 (d, 1H, *J*=11.6, CH₂O). ¹³C NMR (CD₃OD) δ 22.8, 22.8 (COCH₃, C₄), 24.2 (CH₃), 32.3 (C₃), 63.1 (CH₂O), 65.3 (C₁), 76.6 (C₂), 173.3 (CO). ESI⁺ (*m*/ *z*)=174.3. Anal. Calcd for C₈H₁₅NO₃: C, 55.47; H, 8.73; N, 8.09. Found: C, 55.49; H, 8.75; N, 8.07. To a solution of *cis*-**6** (90 mg, 0.22 mmol) in dry THF (8 mL), TBAF (0.22 mL, 1 M in THF) was added and the mixture was stirred for 3 h at 0 °C. Then, the reaction was evaporated in vacuo and the residue was purified by silica gel column chromatography (MeOH/ethyl acetate, 5:95) to obtain the product *cis*-**8** as a colourless oil (30 mg, 80%). ESI⁺ (*m*/*z*)=174.3. ¹H NMR (CDCl₃) δ 1.40 (s, 3H, CH₃), 1.84–1.88 (m, 2H, H₃, H₄), 2.00–2.07 (m, 5H, COCH₃, H₃, H₄), 3.67 (d, 1H, *J*=11.2, CH₂O), 3.86 (d, 1H, *J*=11.2, CH₂O), 6.93 (br s, 1H, NH). ¹³C NMR (CDCl₃) δ 23.4, 23.4 (CH₃, C₄), 23.8 (COCH₃), 32.2 (C₃), 64.2 (C₁), 64.7 (CH₂O), 74.6 (C₂), 172.2 (CO). Anal. Calcd for C₈H₁₅NO₃: C, 55.47; H, 8.73; N, 8.09. Found: C, 55.44; H, 8.72; N, 8.11.

4.1.9. trans-N-(1-(Hydroxymethyl)-2-(methoxymethoxy)-2-methylcyclobutyl)acetamide (trans-**9**)

To a solution of *trans*-**7** (160 mg, 0.35 mmol) in dry THF (15 mL), TBAF (0.35 mL, 1 M in THF) was added and the mixture was stirred for 3 h at 0 °C. Then, the solution was diluted with ethyl acetate (15 mL) and washed with a saturated NH₄Cl solution (20 mL). The organic layer was dried, concentrated in vacuo, and purified by silica gel column chromatography (MeOH/ethyl acetate, 1:9), to give the compound *trans*-**9** as a colourless oil (50 mg, 70%). ESI⁺ (*m*/*z*)=218.4. ¹H NMR (CDCl₃) δ 1.42 (s, 3H, CH₃), 1.83–1.93 (m, 2H, H₃, H₄), 1.96 (s, 3H, COCH₃), 2.09 (dd, 1H, *J*=10.1, *J*=20.2, H₃), 2.19 (dd, 1H, *J*=10.1, *J*=20.3, H₄), 3.34 (s, 3H, OCH₃), 3.76 (d, 1H, *J*=11.7, CH₂OH), 4.07 (d, 1H, *J*=11.7, CH₂OH), 4.62 (d, 1H, *J*=6.9, CH₂OCH₃), 4.67 (d, 1H, *J*=6.9, CH₂OCH₃), 6.18 (br s, 1H, NH). ¹³C NMR (CDCl₃) δ 20.7 (CH₃), 22.2 (C₄), 23.9 (COCH₃), 29.8 (C₃), 55.6 (OCH₃), 63.6 (C₁), 65.5 (CH₂OH), 81.4 (C₂), 91.8 (CH₂OCH₃), 170.5 (CO). Anal. Calcd for C₁₀H₁₉NO₄: C, 55.28; H, 8.81; N, 6.45. Found: C, 55.32; H, 8.75; N, 6.53.

4.1.10. cis-N-(1-(Hydroxymethyl)-2-(methoxymethoxy)-2-methylcyclobutyl)acetamide (cis-**9**)

To a solution of *cis*-**7** (220 mg, 0.48 mmol) in dry THF (20 mL), TBAF (0.48 mL, 1 M in THF) was added and the mixture was stirred for 3 h at 0 °C. Then, the solution was diluted with ethyl acetate (20 mL) and washed with a saturated solution of NH₄Cl (20 mL). The organic layer was dried, concentrated in vacuo, and purified by silica gel column chromatography (MeOH/ethyl acetate, 1:9), to give the compound *cis*-**9** as a colourless oil (80 mg, 75%). ESI⁺ (*m*/*z*)=218.4. ¹H NMR (CDCl₃) δ 1.39 (s, 3H, CH₃), 1.77–1.90 (m, 2H, H₃, H₄), 2.03 (s, 3H, COCH₃), 2.08–2.21 (m, 2H, H₃, H₄), 3.39 (s, 3H, OCH₃), 3.61 (d, 1H, *J*=11.6, CH₂OH), 3.88 (dd, 1H, *J*=6.5, *J*=11.6, CH₂OH), 4.27 (d, 1H, *J*=3.8, OH), 4.67 (d, 1H, *J*=6.9, *CH*₂OCH₃), 4.71 (d, 1H, *J*=6.9, *CH*₂OCH₃), 6.96 (br s, 1H, NH). ¹³C NMR (CDCl₃) δ 20.4 (CH₃), 22.7 (C₄), 24.0 (COCH₃), 30.5 (C₃), 55.8 (OCH₃), 64.5, 64.8 (C₁, CH₂OH), 79.1 (C₂), 92.1 (*CH*₂OCH₃), 171.5 (CO). Anal. Calcd for C₁₀H₁₉NO₄: C, 55.28; H, 8.81; N, 6.45. Found: C, 55.34; H, 8.84; N, 6.47.

Acknowledgements

We thank the Ministerio de Educación y Ciencia (project CTQ2006-05825/BQU), the Gobierno de La Rioja (doctoral fellowship for M.P.-F.), and the CSIC (F.R. JAE-Doc Program contract). We also thank the referees for their valuable suggestions.

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- 9. The cis/trans ratio was determined by ¹H NMR and HPLC when signal overlapping was observed in the ¹H NMR reaction crude spectra. Thus, the data in entries 2, 3 and 10 in Table 1 correspond to ¹H NMR experiments, whereas for the rest of the entries, both the analyses were carried out. HPLC sampling was performed at a flow rate of 1 mL/min (CH₃CN/H₂O, 60/40) using a Phenomenex[®] column, Luna 5 μ m Cl8(2) 100A, 250×4.6 mm. Injection volume=20 μ L. Detector λ =254 nm. Retention times for *cis*-**6** and *trans*-**6** are 19.6 and 20.4 min, respectively.
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